ANTIMICROBIAL PEPTIDES CAN BE AN ALTERNATIVE TO ANTIBIOTICS



Nabanita Chowdhury* Dr. Biswajit saha**

Department of Microbiology Bijoy Krishna Girls' College, Howrah 5/3, Mahatma Gandhi Rd, Howrah, West Bengal 711101, India

*nabanita199926@gmail.com & ** biswajit.saha1402@gmail.com

Abstract: For the rapid growth of the pathogenic bacteria to the conventional antibiotics leads to determine the urgent need for a search of fundamentally new anti-infective drugs. Antimicrobial peptides (AMPs) of the innate immune system are promising conditions for a role of such novel antibiotics. However, some cytotoxicity of AMPs towards host cells limits their active implementation medicine and forces attempts to design numerous structural analogues of the peptides with optimized properties 1. Antimicrobial peptides are widely distributed throughout the animal and plant kingdoms. These peptides are involved in the direct destruction of various microorganisms like bacteria, fungi, parasites, viruses etc. and also can destruct tumor cells. Antimicrobial activities of AMPs, primarily disrupts membranes, so they have a lower likelihood of inducing drug resistance.

Extensive studies show that net charge, hydrophobicity and amphipathicity are the most important physicochemical and structural determinants endowing AMPs with antimicrobial potency and cell selectivity. Designing AMPs for therapy will need to focus on such factors like their susceptibility to proteolytic degradation and reduction of toxicity to mammalian cells. Strict guidelines pertaining to their use should also be established to prevent or hinder future development of bacterial resistance to such peptides 2. This review summarizes the recent advances in the development of AMPs with respect to characteristics, structure-activity relationship, functions, their usage against microbes, expression, regulation and their applications.

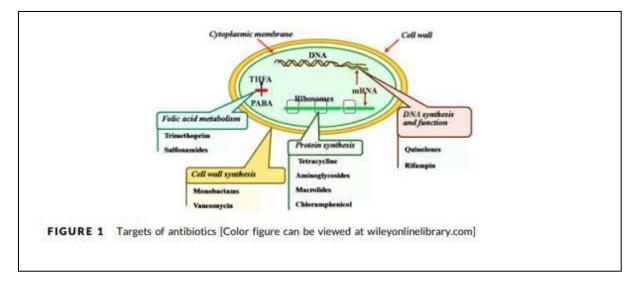
Keywords: Antimicrobial peptide, antimicrobial resistance, therapeutic agents, antibiotics, antibacterial activity, antifungal activity, antiviral peptides, antiparasitic peptides, antitumor activity.

I. INTRODUCTION

Bacterial resistance to various antimicrobial agents has developed as a result of theresponse to their direct exposure. This can be happened, if target protein is altered by mutation or enzymatic activity, or by passing of target protein, or the extrusion of the antimicrobial from the cell

.These adaptations can arise in susceptible bacteria as a result of mutations or through horizontal genetransfer, either within or between genera, primarily employing mobile genetic elements such as plasmids, transposons or integrons 3.Alarmingly, antibiotic-resistant bacteria have been increasingly isolated from patients and animals treated with antibiotics, and this resistance to the most conventional antibiotics has become one of the most pressing global public health concerns

worldwide . Therefore we need to find an alternative antimicrobial strategy 4. Antimicrobial peptides have been described as evolutionary ancient weapons 5. Found in both animal and plant kingdoms, they play a fundamental role in the innate non-specific defence system which confers resistance against infections without prior exposure to foreign pathogens 6 . Antimicrobial peptides are the alternative participants for design of a new antimicrobial agents for their specificity, they have natural antimicrobial properties and a low propensity for the development of bacterial resistance. In addition to rapid and broad-spectrum activities against Gram negative and Gram positive bacteria, fungi, parasites, viruses and tumor cells, AMPs also mediate chemotaxis, apoptosis, immunomoduleatory effects and wound healing 7. The antimicrobial mechanism of AMPs varies from membrane permeabilization to interaction with an array of intracellular target molecules, and differes from that of antibiotics that target specific molecular receptors of pathogens.



The membrane interactions between AMPs and bacteria to acquire resistance against them, which supports the wide application of AMPs in food, medicine and animals for avoiding the overuse of antibiotics. These peptides are stored in granules of phagocytic cells and exert their effects in phage lysosomes or being secreted extracellularly; they are also widely expressed and released at epithelial surfaces and in a site of inflammation 8. They have different amino acid sequences and structures, but most of them are cationic and they can adopt an amphipathic conformation, for this reason they are able to easily interact with the negatively charged components on the surface of the bacterial cells and integrate into the lipid bilayers. The main mechanism of antibacterial action of AMPs is related with their ability to alter membrane permeability and damage its structure 9. Some AMPs are non-membranolytic and penetrate bacterial membranes without disturbing their integrity. They have intracellular targets and interfere with the metabolic processes, including synthesis of the vitally important cell components 10. For this wild-scale multitargeted action it is believed to be one of the reasons for the effectiveness of AMPs towards multidrug -resistance bacterial strains. For this widebeneficial features of AMPs such as broad- spectrum activity, swift and effective bacterial hilling that also complicates the resistance development, would healing promotion, it is thought to be used as a replacement of antibiotics and also in food, animals etc.

1.Discovery of AMPs: Most of the natural antimicrobial peptides are isolated chromatographically from bacteria, plants, fungi and animals. During the first wave ,prior to 1980s, several non-gene encoded antimicrobial peptides were discovered. From 1980s, the second wave started, during this time researchers started their studies over the innate immunity and mechanisms of action of gene-encoded AMPs as potential antimicrobials. Since around 2000, immune modulation properties of AMPs have been reported. In 1922, Sir Alexander Fleming discovered lysozyme, is now recognized as the first AMP. The discovery of lysozyme did not stir up much interest at that time. Lysozyme inhibits bacteria by cleaving saccharides on the cell wall 11. The small protein may be used topically as its size makes it unstable for systemic use 12. Rogeres(1928)1st noticed the ability of nisin to inhibit bacteria, it was the 1st antibiotics. It contains multiple thioether rings. It is the only bacteriocin, which is approved by the USFood and Drug Administration (FDA) as a food preservative. It is used as preservatives of meat and dairy products (<12.5mg/kg food) in over 50 countries. Nisin attacks the cell wall of Gram-positive bacteria like Listeria monocytogens. Nisin also inhibits Gram-negative pathogens such as Escherichia coli and Salmonella spp. When used in combination with chelators or heat treatment 13 .Dubos discovered gramicidin from a soil bacterium Bacillus brevis. Gramicidin A consists of alternating L and D amino acids. The N-terminus of this peptide is formylated 14. It consists of hydrophobic amino acids. Such sequence is viral for the formation of a head-to-head dimer as a membrane channel. It is the 1st peptide-antibiotic which was used clinically as a topical treatment. Gramicidin S isolated from bacteria, used totreat infectious wounds. This small peptide is cyclic, forms peptide bond between the termini. It works both against Gram positive and Gram negative bacteria. Gramicidin S is still used in topical oinments and eye drops 15. Polymixin E (Colistin) is also a bacteriocine is stillused clinically to treat infection caused by Gram negative pathogens. It has a cyclic peptide structure followed by a lipid tail 16. Daptomycin has a similar structure, with a negative charge. It requires the presence of Ca2+ to show its full activity. In 2003 Daptomycin wasapproved by FDA to treat Gram positive bacteria infections. Alamethicin isolated from the fungus Trichoderma viride is also an AMP. It consists of seven alpha-aminoisobutyric acids forming a helical structure. It prevents Gram positive bacteria and fungi. This is perhaps the only AMP with evidence to support to barrel-stone pore in membranes 17. Boman et al. discovered ceeropins(1980s) from the moth Hyalophora Ceeropia. Lehrer and his colleagues identified the first alpha defensins from human neutrophils 18. Subsequently the first beta defensin was discovered from cattle 19. Cyclic thetadefensin was discovered by Tang et al. (1999). All of these defensins contain three pairs of disulfide bonds 20. Due to their small size and stability, there is substantial interest in developing therapeutic uses for theta defensin miniproteins 21.Lucifesin is a defensin, discovered in 2010 from insects. This is probably a key antimicrobial element for maggot therapy.

In summary, all of these peptides mentioned above, is used as antimicrobial agents, also some are under development. In addution, the induction of AMP expression, at a needed sitre and tine, provides a new avenue for antimicrobial development 22.

II NOMENCLATURE OF ANTIMICROBIAL PEPTIDES:

Although various methods are employed for naming the newly identified peptides, there are three mostcommon methods present

i)Source base methods

ii)Peptide base method

iii)Source and peptide combined methods

<u>i)</u> <u>SOURCE BASED METHOD:</u> It is the most common method. In this method either species or genus name is taken.Example : Sesquin is derived from *vigna sesquipedalis*, here the name comes from the species name. Palicourein is derived from *Palicourea condensala*, here the name derived from the genus name.Sometimes peptide name is derived from the common name of the organism eg, termicin from termites. Also abbreviations of animal names are utilized such that bBD-1(bovine beta defensing -1). Other animal abbreviations include PMAP-36 where P indicates pigs. e-CATH-1, e indicates equine. Sometimes, names of organs or tissues are also utilized . eg, Human neutrophil peptide-1(HNP-1) Liver expressed antimicrobial peptide-2 (LEAP-2).

ii) PEPTIDE BASED METHOD: Same AMPs are named according to their peptides' properties. The name of defensin is derived from the word 'defence', implying the functional role of this family of peptides. On the other hand many AMPs are named after their amino acid sequences.Example : Humanhistatins are named like this as they are rich in histidine residues. PR-39 consists of 39 residues which is rich in proline and arginine residues.

<u>iii</u>) **SOURCE AND PEPTIDE COMBINED METHOD :** In some cases, source of organisms and peptide features are combined to assign a unique name of AMPs.For instance,Ib-AMP is abbreviated from Impatiens balsamina antimicrobial peptide. When there are multiple similar peptides, they are named by giving numbers such as Ib-AMP1,Ib-AMP2,Ib-AMP3, Ib-AMP4 etc.

III MAJOR PARAMETERS THAT DETERMINE THE ACTIVITY OF ANTIMICROBIAL PEPTIDES

i) **LENGTH :** Length of the antimicrobial peptides play a major role to determine their activity as at least 7-8 amino acids are required to form amphipathic structures with hydrophilic and hydrophobic faces on opposite sides of a peptide molecule. In the barrel- stave model at least 22 amino acids should be present for alpha-helical AMPs and at least 8 amino acids areneeded for beta-sheet AMPs. Besides the effect of length on the mode of action, it also affect its cytotoxicity. For example: a shortened melttin and a shorter derivative of HP(2-20)exhibited at least 300 times less toxicity to rat erythrocytes, respectively, compared to their original length. Therefore the length of AMP should be taken into consideration when designing a new synthetic peptides with low toxicity.

- ii) **CHARGE:** The net charge of the AMPs is the sum of all charges of ionizable groups of thepeptide. Charge of AMPs varies from positive to negative, and it is one of the most important factor for it's antimicrobial activity. Most of them are positively charged; ranging from +2 to +9, the interaction between AMPs and cell membrane mainly relies an electrostatic attraction. It is generally assumed that cationic AMPs initially interact with negatively charged lipid head groups on the outer surface of the cytoplasmic membrane. The peptide than penetrates the outer leaflet of the cytoplasmic membrane lipid bilayer in an approximately parallel orientation to the bilayer, which leads to the displacement of lipids23. But an excessive charge can affect it's antimicrobial activity. Continuously increasing the number of positive charges may not increases it's activity. Same anionic AMPs are also present, they are rich in Glu and Asp, they participate in the eukaryotic innate immune response. These peptides have not charges ranging from-1 to -2, and they generally require cations for .zn²+, as compactors for biological activity. By changing the net charge of an AMP, its antimicrobial and hemolytic activities can be altered to achieve selective killing of microbes with no or minimized effects on host cells. For eg:, increasing positive net charge of V13 K from +8 to +9 resulted in higher hemolytic activity, while decreasing the net charge to lower than +4 abolished its activity against P. aeroginase 24.
- **HYDROPHOBICITY:** It is the another important factor which influence the activity iii) and selectivity of APMs . Almost 50% of amino acids in primary sequence of natural AMPs are hydrophobic residues 25. In most of the cases increase in hydrophobicity on the positively charged side below a threshold value can increase its antimicrobial activity. On the other hand, decreasing the hydrophobicity can reduce antimicrobial activity. Each and every AMPhas an optimal hydrophobicity, beyond which its activity decreases rapidly. Therefore while designing new synthetic peptides, the hydrophobicity should be selected at around optimal value. Some studies shown that hydrophobicity is also a critical component for selecting therange of target cells of an AMP. By changing the hydrophobicity of an AMP it's target range can be changed. For eg., magainin is an AMP that is only effective against Gram-positive bacteria. However, some synthetic analogs with higher hydrophobicity can also kill some Gram-positive bacteria and eukaryotic cells 26. AMPs with high hydrophobicity can damage the membrane structure, which results in cell lysis or the formation of transient pores and the transport of peptidesinside the cell, this property enables them to interact with intracellular targets 27.
- IV) **Amphipathicity:** It is the another important property of AMPs which ensure the activity and interaction with microbial membranes.
- Fermandes -Vidal et al. showed that amphipathicity is more important than hydrophobicity for binding to microbial membranes.

Because the amphipathicity of AMPs is required for a strong partition into the membrane interface, priority should be given to the amphipathic structure when designing synthetic AMPs for specific target cells.

It is the most important physicochemical and structural parameters for AMPs antimicrobial activity. It results from the segregation of hydrophobic and polar residues on the opposite face of the molecular framework. Some studies suggest that perfect amphipathicity often results in a simultaneous increase in both bactericidal and cytotoxic activity. Wanget al found that imperfectly amphiphilic peptides should better antimicrobial activity than the corresponding perfectly amphipathic peptides.

83

V) <u>**Covalent Bonds:**</u> Covalent bonds have profound effect on AMPs. The modification the covalent bonds should have positive or negative effect on AMPs work means it can increase or reduce it's antimicrobial activity. For eg.,Protegrin missing a disulphide bond becomes inactive against HSV; while adding disulphide bond in Sakacin P resulted in higher antimicrobial activities 28. Another studies shows that addition of a disulphide bond and a trp-trp crosslink to indolicin, increase its stability against protease with no change in antimicrobial activity.

However increase in stability does not always lead to better AMPs. For eg., Houston et al. introduced a covalent bond to form a lactam bridge between Gln and Lysresidues in two alpha-helical AMPs, eg., Cecropin and mellitin. This modification helped AMPs to form more stable alpha-helix structures but decreased the antimicrobial activity of both.

IV STRUCTURE OF AMPS

Primary structure:

<u>Sequence Length:</u> AMPs have variable sequence length ranging from 10-60 amino acid residues shorter AMPs are preferred for their low producing cost, many of them show similar antimicrobial activities to those of longer AMPs, eg., the hexapeptide MP196 (RWRWRW-NH2) shows robust activity against E.coli and S. aureus with an minimum inhibitory concentration(MIC) value of 5microgram/liter

29. Moreover, long chain linear peptides are often more hemolytic and cytotoxic but their N or C- terminal- truncated sequences usually have a lower cytotoxicity but retain robust activity 30. However, peptide with very short lengths shows a decreased tendency to form amphipathic secondary structures, which leads to low antimicrobial potency. So, it can be said that AMPs have a certain threshold length for binding with membranes with a high efficiency and form helical structures.

<u>Amino acid composition:</u> AMPs have mainly two types of amino acid residues, cationic and hydrophobic residues. Naturally occurring AMPs contain Arg; Lys and His amino acid residues and hydrophobic residues are mainly alipathic and aromatic amino acids. The positively charged amino acidresidues of AMPs directly interact with the negatively charged component of bacterial cells. Then, the hydrophobic residues get incorporated into lipid bilayers to mediate membrane permeabilization and disruption, which lead to rapid cell death.

Characteristics of secondary

structures: Alpha-helix:

According to updated database, the proportion of the natural alpha-helical peptides is the highest among AMPs (fig 2), and they are mainly derived from different species, including insects, fish, amphibians, mammalians and plants. Many studies have found that most of these peptides, interacting with membranes are converted to alpha-helical structures. This transformation induces the segregation of hydrophilic/ charged amino acid in space from the hydrophilic residues, which results in an amphipathic structure that is recognized as a prerequisite for AMPs to act on membranes 31.

Sequence analysis of PMAP-36 showed that it has the highest proportion of cataionic amino acids (36%),most frequently at the N- terminus. Structural analysis of PMAP-36 further demonstrate

that this highly cationic sequence adopts a typical amphipathic alpha-helical conformation and random hydrophobic tail. Recent studies have indicated that N-terminal region (alpha-helical domain) of PMAP-36 is the active region 32. Researchers have found that the disruption of amphipathicity increased antimicrobialactivity with very little hemolysis.

According to the helical wheel projection, Ma et al designed a small combinatorial library of Val/ Arg - rich peptides, and the peptide G6 was found to have optimal cell selectivity. G6 has been further demonstrated to reduce peritoneal bacterial counts and increase survival after *Salmonella typhimurium* infection 33.

It was found that the net increase in charge increase antimicrobial activity and decrease hemolysis and the substituition of Gly with Pro significantly reduce toxicity. Sources of some AMPs along with their chemical and biochemical properties are summarized in Table 1.

Beta-sheets: Besides alpha-helixes, beta-sheets are another principal secondary structures of AMPs. Inaqueous solution most of the peptides change their conformation from unstructured forms to beta- sheet structures in a membrane mimetic environment.

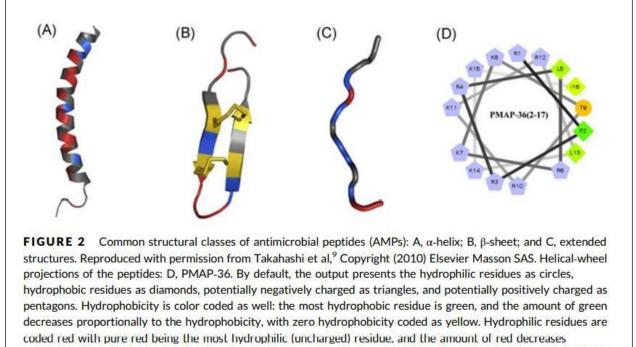
Generally beta-sheet peptides consist of two to ten cys residues, they form disulfide bridges for stabilizing their bioactive conformation. Additionally, peptides that contain disulfide bonds often adopt a cyclic beta-hairpin conformation, such as tachyplesin and protegrin 1 (PG 1) 34. The turn region of PG-1 contains three positively charged Arg residues, and this is considered to be active center of PG-1.

Natural beta- sheet AMPs play important role in innate immune system by killing invading pathogens or regulating immune responses. Defensins have six to eight Cys residues in defined positions, conserved across most multicellular organisms.

Peptides that typically adopt alpha-helix and beta-sheet structures have been designed to improve the prospects for applying AMPs therapeutically and the relationship between structures and bioactivities has been investigated 35. The beta-sheet structure posses greater cell selectivity than that of alpha- helical peptides with equal hydrophobicity and charge. But it is difficult to form robus beta-structure forshort peptides. Studies have shown that several rational designs of synthetic beta-sheet folding peptide amplifies broad-spectrum and highly selective antimicrobial activities. Therefore, the protein folding theory and the common features of natural AMPs provide a basis for the design of beta-sheet AMPs, including:(1)the intrinsic beta-sheet propensities of amino acids in strands and cross-strand interactions across strands, (2) a net positive charge mediating peptide interactions with negatively charged membranes of bacteria; (3) hydrophobic residues providing lipophilic anchors and ultimately including membrane disruption, and (4) the reasonable arrangement of amino acid residues to form a structure with amphipathic characteristics, which segregates cationic and hydrophobic residues to opposite faces of the folded molecule 36. Researchers have found that synthetic hairpin structure of a S.typhimurium strain provided resistance to bacterial infection. Subsequently, a series of symmetric end beta-sheet peptides showed the effect of the type of amino acid on AMP activity.

Extended structures: Most of the extended properties with a high proportion of Pro and Gly residues usually exhibit a linear structure, rather than typical secondary structures. Therefore, extended properties can be divided into Pro and Gly-rich peptides 37. Indolicidin has Pro-rich extended property and it is isolated from mammals, on the other hands apidaecins are isolated from insects. Pro-residues varies from 15-39 residues. The pro-residue is often associated in doublets and triplets with basic residues (Arg and Lys). Generally short Pro-rich peptides exhibit

effective activity against Gram negative bacteria while maintain low antimicrobial activity against Gram positive bacteria. Several Gly-rich AMPshave been isolated from various insects. The size of Gly-rich peptides varies from 8kDa (holotricin) to 30kDa (sarcotoxin II) 38.



proportionally to the hydrophilicity. The potentially charged residues are light blue. Reproduced with permission from Zhu et al,³⁰ Copyright (2014) Elsevier Ltd [Color figure can be viewed at wileyonlinelibrary.com]

| Classes | Representatives | Sequences | Hosts |
|----------|-----------------|--|-----------|
| α-Helix | Melittin | GIGAVLKVLTTGLPALISWIKRKRQQ | Honey bee |
| | Magainin-1 | GIGKFLHSAGKFGKAFVGEIMKS | Frog |
| | LL-37 | LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNLVPRTES | Human |
| β-Sheet | HNP-1 | AC[1]YC[2]RIPAC[3]IAGGRRYGTC[2]IYGGRKWAFC[3]C[1] | Human |
| | HBD-1 | DHYNC[1]VSSGGQC[2]LYSAC[3]PIFTKIQGTC[2]YRGKAKC[1]C[3]K | Human |
| | Protegrin 1 | RGGRLC[1]YC[2]RRRFC[2]VC[1]VGR | Pig |
| Extended | Indolicidin | ILPWKWPWWPWRR | Cow |
| | Tritrpticin | VRRFPWWWPFLRR | Pig |
| | PR-39 | RRRPRPPYLPRPRPPFFPPRLPPRIPPGFPPRFPPRFP | Pig |

V MODE OF ACTION

Different AMPs have different modes of action, some kill cells by disrupting membrane integrity, some by inhibiting proteins, DNA, RNA synthesis or several by interacting with

certain intracellular targets. All AMPs known by late-90s are cationic. However, the concept that AMPs need to be cation was changed later with the discovery of negatively charged AMPs in 1997 39. Eg.

Maximin-H5 is an anionic peptide, isolated from frog skin.

Generally an particular AMPs is active against only one specific class of microorganisms (eg. Bacteria orfungi) . But, there are some exceptions, some AMPs have several mechanisms of action. For eg.

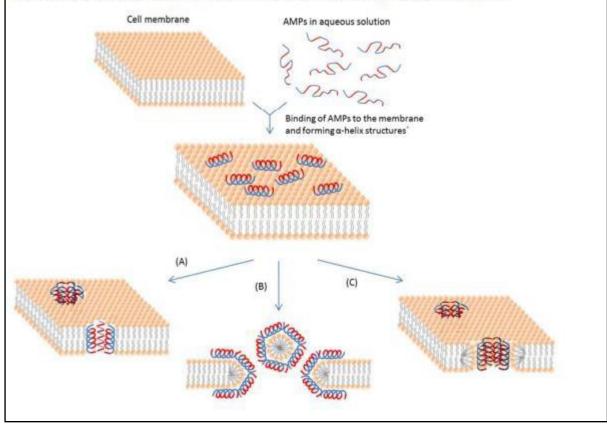
Indolicidin can kill bacteria, fungi and HIV 40. However, it kills *E.coli* by penetrating into the cells and inhibiting DNA synthesis, and it shows anti-HIV activities by inhibiting HIV-integrase 41. Also there are some AMPs which kill different types of cells by the same mode. For eg. PMAP-23 can kill both fungi and parasites by forming pores in their cell membranes 42.

Approximately one third of the proteins of a bacterial cell are associated with the membrane and these proteins have many functions that are critical to the cell including active transport of nutrients, respiration, proton motive force, ATP generation and intercellular communication 43. Without complete cell lysis, with AMP treatment, the function of these proteins can be altered. Therefore, AMPs does not only can disrupt cell membrane but can also inhibit some functional proteins.

<u>Membrane-active AMPs</u>: Most membrane-active AMPs are amphipathic, i.e, they both have hydrophobic and cationic faces. This feature ensures the initial electrostatic interaction with the negatively charged cell membrane and the hydrophobic part helps the AMP molecule to insert into thecell membrane. So, the interaction mainly depends on cationic state and hydrophobicity of the AMP molecule. The major types of membrane-active AMPs and the mechanisms of their actions are summarized in Table 2 and fig. 3.

| Interaction model | Mechanism | References |
|---------------------------------|---|---------------|
| Carpet like (Detergent-like) | The peptide micelle touches the membrane first and coats a small area of the membrane. Then AMP molecules penetrate the lipid bilayer to let pore formation occur leaving holes behind. | [115–117] |
| Membrane thinning | AMPs insert themselves into only one side of the lipid bilayer. It can form a gap between lipid molecules at the chain region. This gap creates a force and pulls the neighboring lipid molecules to fill it. | [118-120] |
| Aggregate | AMPs stick to the membrane parallel to the surface. Then reorientation of AMPs occurs and they insert themselves into the membrane vertically to form sphere-like structures. | [115,121-123] |
| Toroidal pore | AMPs align perpendicularly into the bilayer structure with their hydrophobic regions associated with the center part of the lipid bilayer and their hydrophilic regions facing the pore. | [83,123] |
| Barrel-stave | Staves are formed first parallel to the cell membrane. Then barrels are formed and AMPs are inserted perpendicularly to the plane of the membrane bilayer. | [82,124,125] |

Figure 3 Schematic representation of some action mechanisms of membrane-active AMPs. (A) Barrel-Stave model. AMP molecules insert themselves into the membrane perpendicularly. (B) Carpet model. Small areas of the membrane are coated with AMP molecules with hydrophobic sides facing inward leaving pores behind in the membrane. (C) Toroidal pore model. This model resembles the Barrel-stave model, but AMPs are always in contact with phospholipid head groups of the membrane. The blue color represents the hydrophobic portions of AMPs, while the red color represents the hydrophilic parts of the AMPs.



Intracellularly Active AMPs: Intracellularly active AMPs have been shown to interact with targets inside the cells 44. For example indolicin was shown to bind to DNA with a preferred sequence 45. Some AMPscan inhibit DNA and protein synthesis 46. One example of this is PR-39, an AMP from pig intestines, which kills bacteria in a non-lytic process by acting like a proteolytic agent and stopping protein and DNA synthesis 47. Similar to PR-39, indolicin does not lyse cells directly. It enters the cytoplasm and kills bacterial cells by targeting DNA synthesis 48.

VI BROAD SPECTRUM ACTIVITY:

<u>Antifungal activity:</u> AMPs have strong antifungal activity and it is also useful in addressing fungal infections. AMPs generally lyse the cell and they interfare with fungal cell wall synthesis. Cathelicidin peptides destroyed Candida albicans and Cryptococcus neoformans cells via membrane permeabilization and damage the microorganism.

Vylkova et al. found that human beta-defensin 2 (Hbd-2)(Synthetic) and Hbd-3(synthetic) could destroy *Candida albicans* in an energy – dependent and salt-sensitive manner without causing gross membrane disturbance or lysis 49. Research has shown that LL-37 could reduce the *C.albicans* attachment to abiotic surfaces, oral epidermis and murin urinary bladders by interacting with Yeast carbohydrate and protein cell- wall components, which is of critical importance inprevention *C.albicans* colonization and infection by AMPs 50.

Antiviral peptides: Antiviral AMPs neutralize viruses by interacting in either the viral envelope or the host cell membrane. AMPs can integrate into viral envelopes causing membrane instability, this render the viruses unable to infect host cells. For eg., defensins bind to the viral glycoproteins making herpes simplex viruses (HSV) unable to bind to the surface of the host cells 51. There are some antiviral AMPs which can prevent viral particles from entering host cells by occupying specific receptors on mammalian cells. For eg., heparan sulfate is important for the attachment of HSV viral particles to the host cell surface. The heparan sulfate molecules are negatively charged glycosaminoglycan molecules. Thus, some alpha-helical cationic peptides, eg., lactoferrin, can prevent HSV infections by binding to heparan molecules and blocking virus-receptor interactions 52. Some AMPs can cross the cell membrane and localize in the cytoplasm and changes in the gene expression profile of host cells, which can help host defense system to fight against viruses or can block viral gene expression. For eg., NP-1, prevents Vero and CaSki cell lines from infection by herpes simplex viruses type 2 (HSV-2). This AMPs stops the viruses by preventing the migration of a major viral protein, VP16, into the nucleus. This viral protein is required to form complexes with the host transcriptional factors to induce the expression of immediate early viral genes, which are required for the virus to defeat the first stage cellular response 53. Thus, this AMP does not compete with viral particles to bind to the receptor on cell surface but it prevents cell-to-cell spread of viral particles.

<u>Antibacterial activity:</u> Antibacterial AMPs are the most studied AMPs. Most of them are cationic AMPs. These AMPs target bacterial cell membranes and cause disintegration of the lipid bilayers. Themajority of these AMPs are also amphipathic with both hydrophilic and hydrophobic domains. Such structures provide AMPs the capability to bind to lipid components (hydrophobic region) and phospholipid groups (hydrophilic region) 54.

Researchers have demonstrated that some AMPs at low concentration can kill bacteria by inhibiting some important pathways inside the cell. For eg., buforin II can diffuse into cells and bind to DNA and RNA without damaging the cell membrane 55. Drosocin, pyrrhocoricin, and apidaecin are other examples of such AMPs. It is generally hypothesized that three main mechanisms could account for peptide permeation of the membrane of the target cell, including "barrel-stave model", "carpet model" and "toroidal-pore model" 56. In the "barrel-stave model", the attached peptides aggregate to form a bundle with a central lumen and insert into the hydrophobic coreof the membrane forming a trans-membrane pore. The "carpet model" suggests that AMPs bind onto the phospholipid head covering the surface of membranes in a carpet-like manner and disrupt the bilayer curvature like a detergent beyond a threshold concentration of membrane-bound peptide 57.

The "toroidal-pore model" involves aggregation of peptide helices into the membrane, inducing the lipid monolayers to bend continuously through the pore so that both the inserted peptides and the lipid headgroups line the water core 58.

Antiparasitic peptides: The first antiparasitic AMP is magainin. It can kill Paramecium caudatum. Later, a synthetic peptide was developed against Leishmania parasite 59. Cathelicidin is an antiparasitic AMP, which is able to kill *Caernohabditis elegans* by forming pores in the cell membrane, the mode of action of antiparasitic peptides is the same as other AMPs. They kill cells by directly interacting with cellmembrane.

<u>Antitumor activity:</u> Cancer was the second leading cause of death, after heart disease, in the United States in 2019. Therefore, development of antitumor drug is needed. Some cationic AMPs exhibit cytotoxic activity against tumor cells. AMPs mainly act on target cell membranes via a non-receptor- mediated pathway, for which it is more difficult for tumor cells to develop resistance compared to conventional chemotherapeutic agents 60. For example, the antitumor mechanism of B1 and its analogs involves three steps: cell membrane disruption resulting from changes in membrane permeability; penetration of the cytoplasm after membrane disruption; disruption of mitochondrial membranes and release of cytochrome C 61.

AMPs with ideal antitumor activity should meet at least three conditions: (1) high net positive charge. The existence of the high net positive charge of AMPs contributes to electrostatic attraction between the negatively charged components (such as phosphatidylserine) of tumor cells and the positively charged AMPs 62. (2) High structural flexibility. Theflexibly of AMPs allows for changes in conformation in different environments (aqueous and membrane-mimic environments), which allows them to traverse the phospholipid layer of tumour cells. (3) High oligomerization. AMPs should be easily clustered on the membrane surface of a tumour cell so that they can form a pore on the tumour cell membrane.

VII APPLICATION OF ANTIMICROBIAL PEPTIDES:

In Food: In food industries natural AMPs are needed without toxicity, to preserve foods. In recent years, much attention has been focused on application of AMPs as an alternative to control undesirable microbial growth on foodstuffs. For eg, Nisin , a polycyclic peptide with 34 amino acids residues derived from Lactococcus lactis , is usually used in processed cheese, meats and beverages.

Ple/ PVA fiber, an AMP Ple (pleurocidin, a novel AMP with 25 amino acid, derived from the skin-secreted mucous of the winter flounder) incorporated into ultrafine PVA fiber mats via electrospinning technology, was demonstrated to be successfully applied in apple cider , with efficient inhibition activity against *E.coli*.

In medicine: AMPs also use in drug therapy in the clinics. Eg, Lucifensin and Lucifensin II, two insect defensins , can heal wound, especially in patients with impaired healing due to underlying disorders(eg.Diabetes). This procedure is known as debridment therapy. Pexiganan, a 22-amino-acid membrane disruptor analog of the Xenopus peptide magainin, has been clinically proven to replace ofloxacin in thetreatment of diabetic foot ulcers as early as 1996, and may avoid the selection of resistant bacteria thatcan develop after oral systemic antibiotic therapy; however, the peptide was not approved by the Food and Drug Administration in 1999 even after completion of a phase III trial. Currently the phase III clinicaltrials of this peptide are being conducted again to treat mildly diabetic foot infection by Dipexium Pharmaceuticals 63. Omiganan is an indolicidin analog. It has a broad-spectrum antibacterial activity. It

has completed phase III clinical trials for catheter infections and rosacea, completed phase II for the application of omiganan in patients with vulvar intraepithelial neoplasia atopic dermatitis and acne vulgaris. POL7080 as an antimicrobial peptidomimetic specifically targets P. aeruginosa at the nanomolar level via non-membrane- disrupting activity. In addition to antibacterial activity, some AMPshave also been shown to have immune-promoting effects, such as PXL01, which is derived from humanlactoferricin and been assessed for its efficacy, safety and handling in patients with flexor tendon injuries in phase II trials 64.

In animals: It has been reported that several AMPs added in the diet have beneficial effects, including body weight, the average daily gain, nutrient digestibility and intestinal morphology as well as effects on intestinal and fecal microflora. AMPs can also improve animal performance, nutrient digestibility and support normal intestinal morphology and function. Some reports also demonstrated that AMPs can protect piglets from challenge with the mycotoxin deoxynivalenol (DON)201 and repair the intestinal injury induced by DON 65. Moreover, AMPs enhance intestinal barrier function and improve microbiota composition in the intestines of weaned piglets and reduce rates of diarrhea in them.

TABLE 3 Selected antimicrobial peptides (AMPs) in clinical phase of development

| AMPs | Description | Condition or disease | Administration | Phase | Status company | Clinical trial identifier if availab |
|-----------------------|---|---|---|---|--|---|
| Pexiganan (MSI-78) | Analog of magainin | Diabetic foot infection | Topical | Phase 3-C | MacroChem Corporation Dipexium Pharmaceuticals, Inc. | NCT00563433 NCT00563394 NCT01594762 NCT01590758 |
| Omiganan | Derived from indolicidin | Catheter infections Atopic dermatitis Rosacea Vulvar intraepithelial neoplasia Acne vulgaris | Topical | Phase 3-C Phase 2-C Phase 3-C Phase 2-C Phase 2-C | Mallinckrodt Cutanea Life Sciences, Inc. | NCT00231153 NCT03091426 NCT02576847 NCT02596074 NCT02571998 |
| Lytixar (LTX-109) | Synthetic antimicrobial peptidomimetic | Gram-positive, skin infections Mild Eczema/Dermatoses Atopic Dermatitis Nasal carriers MRSA Non-bullous Impetigo | Topical | Phase 2-C Phase 1/2-C Phase 2-C | Lytix Biopharma AS | NCT01223222 NCT01158235 NCT01803035 |
| Surotomycin | Cyclic lipopeptide | Clostridium difficile-associated diarrhea (CDAD) | oral | Phase 1-C | Merck Sharp & Dolyme Corp. | NCT02835118 NCT02835105 |
| Novexatin (NP-213) | Cyclic peptide | Onychomycosis | Topical | Phase 2-C | NovaBiotics | 197 |
| Ш-37 | Host-defense peptide | Melanoma | Intratumorally in cutaneous or subcutaneous tumors | Phase 1/2-A | M.D. Anderson Cancer Center | NCT02225366 |
| PXL01 | Derived from lactoferricin | Surgical adhesions | Hyaluronic acid- based hydrogel | Phase 2-C | Pergamum AB | NCT01022242 |
| lseganan (IB-367) | Derived from protegrin 1 | Oral mucositis in head and neck cancer. | Oral rinse | Phase 3-U | National Cancer Institute (NCI) | NCT00022373 |
| PAC-113 | Derived from histatin 3 | Oral candidiasis | Mouth rinse | Phase 2-C | Pacgen Biopharmaceuticals Corporation | NCT00659971 |

Table 3 continued

| AMPs | Description | Condition or disease | Administration | Phase | Status company | Clinical trial identifier if available |
|----------------------------|--|--|--|------------------------|--|---|
| Dalbavancin | Lipoglycopeptide | Bone infection Osteomyelitis Septic arthritis Joint infection Prosthetic joint infection | Intravenously | Phase 4-R | Infectious Diseases Physicians, Inc | NCT03426761 |
| | | Infectious Peritonitis | Intravenously | Phase 4-A | University of Colorado, Denver | NCT02940730 |
| | | Methicillin-resistant Staphylococcus Aureus skin infections | Intravenous administration in children | Phase 3-R | Durata Therapeutics Inc | NCT02814916 |
| SGX942 | 5-amino acid peptide | Oral mucositis in head and neck cancer | Intravenously | Phase 3-R | Soligenix | NCT03237325 |
| OP-145 | Derived from LL-37 | Chronic otitis media | Eardrops | Phase 2-C | Leiden University, The Netherlands | ISRCTN84220089 |
| Brilacidin (PMX- 30063) | Defensin mimetic | Bacterial skin infection Mucositis in head and neck Neoplasms | Intravenously Oral Rinse | Phase 2-C Phase 2-C | Cellceutix Corporation Innovation Pharmaceuticals, Inc | NCT02052388 NCT02324335 |
| POL7080 | Peptidomimetic | Renal impairment Ventilator-associated pneumonia aeruginosa | Intravenously | Phase 1-C Phase 2-C | Polyphor Ltd | NCT02110459 NCT02096328 |
| | | Healthy; Synergism with amikacin | | Phase 1-C | | NCT02897869 |
| AP-214 | Derivative from HDP | Prevention of (acute) kidney injury after cardiac surgery | Intravenously | Phase 2-C | Action Pharma A/S | NCT01256372 |
| | | Prevention of kidney injury after thoracic aortic aneurysm repair | | Phase 2-C | | NCT00903604 |
| CD-NP | Chimeric 37-mer derived from combination of two natriuretic peptides | Acute decompensated heart failure | Infusions | Phase 2-C | Nile Therapeutics | NCT00839007 |
| Ghrelin | Endogenous host-defense peptide | Chronic respiratory infection | Intravenously | Phase 2-C | University of Miyazaki, Japan; | JPRN- UMIN000002599 |
| | | Airway inflammation | | Phase 2-C | | JPRN- UMIN000001598 |

Abbreviations: A, active, not recruiting; C, completed; HDP, host-defense peptide; R, recruiting; U, unknown.

Table 4 Report on application of antimicrobial peptides (AMPs) in animal

| Animals | AMPs | Treatments/Doses | Effects |
|-----------------------------|--|--|--|
| Weanling piglets | CAP | Basal diet with 4 ppm deoxynivalenol and 4% CAP | Attenuating the metabolic disturbances in amino acid, lipid, and energy metabolism induced by DON; Improving intestinal morphology, intestinal epithelial cell proliferation and protein synthesis; Improving fees efficiency, Immune function, and antioxidation capacity, alleviating organ damage |
| Weanling, piglets | SyntheticAMP- A3 ⁶ and AMP-P5 ⁵ | Basal diet with 60 mg/kg AMP-A3 and basal diet with 60 mg/kg AMP-P5 Basal diet with 40 and 60 mg AMP-P5/kg diet Basal diet with 0, 60 and 90 mg AMP-A3/kg diet | Improving the performance, nutrient digestibility, intestinal morphology and reducin pathogenic bacteria |
| Weaned piglets | Cecropin AD ^d | Basal diet with 400 mg/kg cecropin AD and piglets were orally challenged with E. coli K88 | Increasing immune status and nitrogen and energy retention as well as reducing intestinal pathogens |
| Weanling piglets | Recombinant Lactoferrampin- lactoferricin | Basal diet with 0.1 g Lactoferrampin- lactoferricin and 0.1 g chlortetracycline/kg diet | Improving performance and affecting serum parameters |
| Weanling piglets | cipB-LFC-LFA | Basal diet with no addition, 100 mg cipB, 100 mg cipB-LFC-LFA/ kg diet | Improving performance, the regulation of immuni- function and the absorption of Fe, reducin the incidence of diarrhes |
| Weanling piglets | Colicin E1 | Basal diets with0, 11, or 16.5 mg Colicin E1/kg and piglets were orally inoculated with £. coli | Improving the performance and reducing the incidence of postweaning diarrhea |
| Weaned female piglets | Lactoferrin | Basal diet with 1.0 g/kg lactoferrin | Increasing ADG, efficiency of gain, intestinal villus height and relative abundance of mRNA for PR-39 and protegrin 1 |
| Indigenous male chickens | Cecropin AD- Asn (CADN) | Basal diets with a CADN liquid sample at 0, 2, 4, 6, and 8 ml/kg | Increasing nutrient utilization, enhancing intestinal villus heights, decreasing aerobic bacterial counts |

| | (Conti | nued) | | | |
|---------------------------------|----------|-----------------------------|--|--|--------------------------|
| Animals | | AMPs | Treatments/Doses | Effects | References |
| Arbor Ac broiler chickens | | Pig AMP (PMAP) ^e | Basal diet with PAMP at 150 and 200 mg/kg | Improving the performance, the intestinal mucosal immunity, and increasing the intestinal ability to absorb nutrients | Bao et al ²¹² |
| Abbreviatio | ons: AD, | atopic dermatitis; CA | P, composite antimicrobial pe | otides; cipB-LFC-LFA, cipB-lac | toferricin-lactofer- |

VIII PERSPECTIVE AND CONCLUSIONS

Antifungal, antibacterial, antiviral, antiparasitic properties of AMPs promising alternatives to conventional antibiotics. But, there is a limiting factor, that is it has low in vivostability. To extent the half-life of AMPs, AMP mimics with an amphiphilic studies have made great advances in recent days, but further studies are needed. Though many researches have been done on AMPs, infection control by AMP is still hindered by some factors including low specificity, high manufacture cost, potential toxicity to animal cells etc.

In this review we have discussed about various applications of

AMPs, it's mechanism of action, it's various types and it's broad- spectrum activity etc.

IX REFERENCE

[1] Cirioni O, Silvestri C, Ghiselli R, Orlando F, Riva A, Mocchegiani F, Chiodi L, Castelletti S, Gabrielli E, Saba V, Scalise G. Protective effects of the combination of α -helical antimicrobial peptides and rifampicin in three rat models of Pseudomonas aeruginosa infection. Journal of antimicrobial chemotherapy. 2008 Dec 1;62(6):1332-8.

[2] Baltzer SA, Brown MH. Antimicrobial peptides-promising alternatives to conventional antibiotics. Microbial Physiology. 2011;20(4):228-35.

[3] Czaplewski L, Bax R, Clokie M, et al. Alternatives to antibiotics-a pipeline portfolio review. Lancet Infect Dis. 2016,

[4] Zasloff M. Antimicrobial peptides of multicellular organisms. nature. 2002 Jan;415(6870):389-95.

[5] Baltzer SA, Brown MH. Antimicrobial peptides-promising alternatives to conventional antibiotics. Microbial Physiology. 2011;20(4):228-35.

[6] Zhong G, Cheng J, Liang ZC, Xu L, Lou W, Bao C, Ong ZY, Dong H, Yang YY, Fan W. Short synthetic β-sheet antimicrobial peptides for the treatment of multidrug-resistant pseudomonas aeruginosa burn wound infections. Advanced healthcare materials. 2017 Apr;6(7):1601134.

[7] Gallo RL, Hooper LV. Epithelial antimicrobial defence of the skin and intestine. Nature Reviews Immunology. 2012 Jul;12(7):503-16.

[8] Hancock RE, Rozek A. Role of membranes in the activities of antimicrobial cationic peptides. FEMS microbiology letters. 2002 Jan 1;206(2):143-9.

[9]Brogden KA. Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria?. Nature reviews microbiology. 2005 Mar;3(3):238-50.

[10] Wang G, editor. Antimicrobial peptides: discovery, design and novel therapeutic strategies. Cabi; 2017 Sep 1.

[11] Berman HM, Westbrook J, Feng Z, Gilliland G, Bhat TN, Weissig H, Shindyalov IN, Bourne PE. The protein data bank. Nucleic acids research. 2000 Jan 1;28(1):235-42.

[12]Gharsallaoui A, Oulahal N, Joly C, Degraeve P. Nisin as a food preservative: part 1: physicochemical properties, antimicrobial activity, and main uses. Critical reviews in food science and nutrition. 2016 Jun 10;56(8):1262-74.

[13] Wang G, editor. Antimicrobial peptides: discovery, design and novel therapeutic strategies. Cabi; 2017 Sep 1.

[14]Greenwood P. Prevention and intervention programs for juvenile offenders. The future of Children. 2008 Oct 1:185-210.

[15]Stansly PG, Schlosser M. Studies on polymyxin: isolation and identification of Bacillus polymyxa and differentiation of polymyxin from certain known antibiotics. Journal of bacteriology. 1947 Nov;54(5):549-56.

[16]Lang S, Bartl-Pokorny KD, Pokorny FB, Garrido D, Mani N, Fox-Boyer AV, Zhang D, Marschik PB. Canonical babbling: A marker for earlier identification of late detected developmental disorders?. Current Developmental Disorders Reports. 2019 Sep;6(3):111-8.

[17]Ganz T, Selsted ME, Szklarek D, Harwig SS, Daher K, Bainton DF, Lehrer RI. Defensins. Natural peptide antibiotics of human neutrophils. The Journal of clinical investigation. 1985 Oct 1;76(4):1427-35.

[18]Guller S, Markiewicz L, Wozniak RO, Burnham JM, Wang EY, Kaplan PA, Lockwood CJ. Developmental regulation of glucocorticoid-mediated effects on extracellular matrix protein expression in the human placenta. Endocrinology. 1994 May 1;134(5):2064-71.

[19]Wang G, editor. Antimicrobial peptides: discovery, design and novel therapeutic strategies. Cabi; 2017 Sep 1.

[20].Conibear AC, Craik DJ. The chemistry and biology of theta defensins. Angewandte Chemie International Edition. 2014 Sep 26;53(40):10612-23.

[21]Bi, Z., Liang, X., Xu, A., Wang, L., Shi, X., Zhao, W., Ma, J., Guo, X., Zhang, X., Zhang, J. and Ren, J., 2014. Peer Reviewed: Hypertension Prevalence, Awareness, Treatment, and Control and Sodium Intake in Shandong Province, China: Baseline Results From Shandong–Ministry of Health Action on Salt Reduction and Hypertension (SMASH), 2011. *Preventing Chronic Disease*, *11*.

[22] Zhu X, Dong N, Wang Z, Ma Z, Zhang L, Ma Q, Shan A. Design of imperfectly amphipathic α -helical antimicrobial peptides with enhanced cell selectivity. Acta biomaterialia. 2014 Jan 1;10(1):244-57.

[23] Jiang Z, Vasil AI, Hale JD, Hancock RE, Vasil ML, Hodges RS. Effects of net charge and the number of positively charged residues on the biological activity of amphipathic α -helical cationic antimicrobial peptides. Peptide Science. 2008;90(3):369-83.

[24]Tossi A, Sandri L, Giangaspero A. Amphipathic, α-helical antimicrobial peptides. Peptide Science. 2000;55(1):4-30.

[25] Dathe M, Wieprecht T, Nikolenko H, Handel L, Maloy WL, MacDonald DL, Beyermann M, Bienert M. Hydrophobicity, hydrophobic moment and angle subtended by charged residues modulate antibacterial and haemolytic activity of amphipathic helical peptides. FEBS letters. 1997 Feb 17;403(2):208-12.

[26] Pasupuleti M, Schmidtchen A, Malmsten M. Antimicrobial peptides: key components of the innate immune system. Critical reviews in biotechnology. 2012 Jun 1;32(2):143-71.

[27] Johnsen L, Fimland G, Nissen-Meyer J. The C-terminal domain of pediocin-like antimicrobial peptides (class IIa bacteriocins) is involved in specific recognition of the C-terminal part of cognate immunity proteins and in determining the antimicrobial spectrum. Journal of Biological Chemistry. 2005 Mar 11;280(10):9243-50.

[28] Domalaon R, Idowu T, Zhanel GG, Schweizer F. Antibiotic hybrids: the next generation of agents and adjuvants against Gram-negative pathogens?. Clinical microbiology reviews. 2018 Mar 14;31(2):e00077-17.

[29].Luo Y, McLean DT, Linden GJ, McAuley DF, McMullan R, Lundy FT. The naturally occurring host defense peptide, LL-37, and its truncated mimetics KE-18 and KR-12 have selected biocidal and antibiofilm activities against Candida albicans, Staphylococcus aureus, and Escherichia coli in vitro. Frontiers in microbiology. 2017 Mar 31;8:544.

[30] Takahashi D, Shukla SK, Prakash O, Zhang G. Structural determinants of host defense peptides for antimicrobial activity and target cell selectivity. Biochimie. 2010 Sep 1;92(9):1236-41.

[31]Ablikim M, Bai JZ, Ban Y, Bian JG, Bugg DV, Cai X, Chang JF, Chen HF, Chen HS, Chen HX, Chen JC. The σ pole in J/ $\psi \rightarrow \omega \pi^+ \pi^-$. Physics Letters B. 2004 Sep 30;598(3-4):149-58.

[32] Ma QQ, Dong N, Shan AS, Wang L, Hu WN, Sun WY. Biochemical property and In vivo efficacies of novel Val/Arg-rich antimicrobial peptide. Protein and Peptide Letters. 2012 Nov 1;19(11):1144-8.

[33]Bhunia A, Mohanram H, Domadia PN, Torres J, Bhattacharjya S. Designed β-Boomerang Antiendotoxic and Antimicrobial Peptides: STRUCTURES AND ACTIVITIES IN LIPOPOLYSACCHARIDE*. Journal of Biological Chemistry. 2009 Aug 14;284(33):21991-2004.

[34] Hyper-Kamiokande proto-Collaboration, Abe K, Abe K, Ahn SH, Aihara H, Aimi A, Akutsu R, Andreopoulos C, Anghel I, Anthony LH, Antonova M. Physics potentials with the second Hyper-Kamiokande detector in Korea. Progress of Theoretical and Experimental Physics. 2018 Jun;2018(6):063C01.

[35] Zhu X, Dong N, Wang Z, Ma Z, Zhang L, Ma Q, Shan A. Design of imperfectly amphipathic α -helical antimicrobial peptides with enhanced cell selectivity. Acta biomaterialia. 2014 Jan 1;10(1):244-57.

[36] Bulet P, Hetru C, Dimarcq JL, Hoffmann D. Antimicrobial peptides in insects; structure and function. Developmental & Comparative Immunology. 1999 Jun 1;23(4-5):329-44.

[37] Krizsan A, Volke D, Weinert S, Sträter N, Knappe D, Hoffmann R. Insect-derived proline-rich antimicrobial peptides kill bacteria by inhibiting bacterial protein translation at the 70 S ribosome. Angewandte Chemie International Edition. 2014 Nov 3;53(45):12236-9.

[38]Brogden KA, Ackermann M, Huttner KM. Detection of anionic antimicrobial peptides in ovine bronchoalveolar lavage fluid and respiratory epithelium. Infection and immunity. 1998 Dec 1;66(12):5948-54.

[39] Robinson Jr WE, McDougall B, Tran D, Selsted ME. Anti-HIV-1 activity of indolicidin, an antimicrobial peptide from neutrophils. Journal of leukocyte biology. 1998 Jan;63(1):94-100.

[40] Krajewski K, Marchand C, Long YQ, Pommier Y, Roller PP. Synthesis and HIV-1 integrase inhibitory activity of dimeric and tetrameric analogs of indolicidin. Bioorganic & medicinal chemistry letters. 2004 Nov 15;14(22):5595-8.

[41]Lee DG, Kim PI, Park Y, Woo ER, Choi JS, Choi CH, Hahm KS. Design of novel peptide analogs with potent fungicidal activity, based on PMAP-23 antimicrobial peptide isolated from porcine myeloid. Biochemical and biophysical research communications. 2002 Apr 26;293(1):231-8.

[42]Zhang YM, Rock CO. Membrane lipid homeostasis in bacteria. Nature Reviews Microbiology. 2008 Mar;6(3):222-33.

[43]Choi KY, Mookherjee N. Multiple immune-modulatory functions of cathelicidin host defense peptides. Frontiers in immunology. 2012 Jun 11;3:149.

[44] Sieprawska-Lupa M, Mydel P, Krawczyk K, Wójcik K, Puklo M, Lupa B, Suder P, Silberring J, Reed M, Pohl J, Shafer W. Degradation of human antimicrobial peptide LL-37 by Staphylococcus aureus-derived proteinases. Antimicrobial agents and chemotherapy. 2004 Dec;48(12):4673-9.

[45] Hilpert K, McLeod B, Yu J, Elliott MR, Rautenbach M, Ruden S, Bürck J, Muhle-Goll C, Ulrich AS, Keller S, Hancock RE. Short cationic antimicrobial peptides interact with ATP. Antimicrobial agents and chemotherapy. 2010 Oct;54(10):4480-3.

[46] Andersson M, Gunne H, Agerberth B, Boman A, Bergman T, Sillard R, Jörnvall H, Mutt V, Olsson B, Wigzell H. NK-lysin, a novel effector peptide of cytotoxic T and NK cells. Structure and cDNA cloning of the porcine form, induction by interleukin 2, antibacterial and antitumour activity. The EMBO journal. 1995 Apr;14(8):1615-25.

[47] Subbalakshmi C, Sitaram N. Mechanism of antimicrobial action of indolicidin. FEMS microbiology letters. 1998 Mar 1;160(1):91-6.

[48] Vylkova S, Nayyar N, Li W, Edgerton M. Human β -defensins kill Candida albicans in an energy-dependent and salt-sensitive manner without causing membrane disruption. Antimicrobial agents and chemotherapy. 2007 Jan;51(1):154-61.

[49]Tsai PW, Yang CY, Chang HT, Lan CY. Human antimicrobial peptide LL-37 inhibits adhesion of Candida albicans by interacting with yeast cell-wall carbohydrates. PloS one. 2011 Mar 14;6(3):e17755.

[50] Yasin B, Wang W, Pang M, Cheshenko N, Hong T, Waring AJ, Herold BC, Wagar EA, Lehrer RI. θ defensins protect cells from infection by herpes simplex virus by inhibiting viral adhesion and entry. Journal of virology. 2004 May 15;78(10):5147-56.

[51] Andersen JH, Jenssen H, Sandvik K, Gutteberg TJ. Anti-HSV activity of lactoferrin and lactoferricin is dependent on the presence of heparan sulphate at the cell surface. Journal of medical virology. 2004 Oct;74(2):262-71.

[52] Sinha S, Cheshenko N, Lehrer RI, Herold BC. NP-1, a rabbit α -defensin, prevents the entry and intercellular spread of herpes simplex virus type 2. Antimicrobial agents and chemotherapy. 2003 Feb;47(2):494-500.

[53] Jenssen H, Hamill P, Hancock RE. Peptide antimicrobial agents. Clinical microbiology reviews. 2006 Jul;19(3):491-511.

[54] Park CB, Kim HS, Kim SC. Mechanism of action of the antimicrobial peptide buforin II: buforin II kills microorganisms by penetrating the cell membrane and inhibiting cellular functions. Biochemical and biophysical research communications. 1998 Mar 6;244(1):253-7.

[56] Yang L, Harroun TA, Weiss TM, Ding L, Huang HW. Barrel-stave model or toroidal model? A case study on melittin pores. Biophysical journal. 2001 Sep 1;81(3):1475-85.

[57]. Shai Y. Mechanism of the binding, insertion and destabilization of phospholipid bilayer membranes by α -helical antimicrobial and cell non-selective membrane-lytic peptides. Biochimica et Biophysica Acta (BBA)-Biomembranes. 1999 Dec 15;1462(1-2):55-70.

[58]Wang S, A Thacker P, Watford M, Qiao S. Functions of antimicrobial peptides in gut homeostasis. Current Protein and Peptide Science. 2015 Nov 1;16(7):582-91.

[59]Alberola J, Rodriguez A, Francino O, Roura X, Rivas L, Andreu D. Safety and efficacy of antimicrobial peptides against naturally acquired leishmaniasis. Antimicrobial agents and chemotherapy. 2004 Feb;48(2):641-3.

[60]Mader JS, Hoskin DW. Cationic antimicrobial peptides as novel cytotoxic agents for cancer treatment. Expert opinion on investigational drugs. 2006 Aug 1;15(8):933-46.

[61] Deng X, Qiu Q, Yang B, Wang X, Huang W, Qian H. Design, synthesis and biological evaluation of novel peptides with anti-cancer and drug resistance-reversing activities. European Journal of Medicinal Chemistry. 2015 Jan 7;89:540-8.

[62]Harris F, Dennison SR, Singh J, Phoenix DA. On the selectivity and efficacy of defense peptides with respect to cancer cells. Medicinal research reviews. 2013 Jan;33(1):190-234.

[63] Greber KE, Dawgul M, Kamysz W, Sawicki W. Cationic net charge and counter ion type as antimicrobial activity determinant factors of short lipopeptides. Frontiers in microbiology. 2017 Feb 1;8:123.

[64] Mahlapuu M, Håkansson J, Ringstad L, Björn C. Antimicrobial peptides: an emerging category of therapeutic agents. Frontiers in cellular and infection microbiology. 2016 Dec 27;6:194.

[65] Xiao H, Wu MM, Tan BE, Yin YL, Li TJ, Xiao DF, Li L. Effects of composite antimicrobial peptides in weanling piglets challenged with deoxynivalenol: I. Growth performance, immune function, and antioxidation capacity. Journal of animal science. 2013 Oct 1;91(10):4772-80.